

Inside the Diabetic Brain: Role of Different Players Involved in Cognitive Decline

Joana M. Gaspar,^{*,†,‡,§} Filipa I. Baptista,^{‡,||} M. Paula Macedo,^{†,§} and António F. Ambrósio^{‡,||,⊥}

[†]CEDOC, Chronic Diseases Research Centre, NOVA Medical School/Faculdade de Ciências Médicas, Universidade Nova de Lisboa, Edifício CEDOC - IIRua Câmara Pestana no. 6, 6A e 6B, 1150-082 Lisboa, Portugal

[‡]Institute for Biomedical Imaging and Life Sciences (IBILI), Faculty of Medicine, University of Coimbra, 3000-548 Coimbra, Portugal

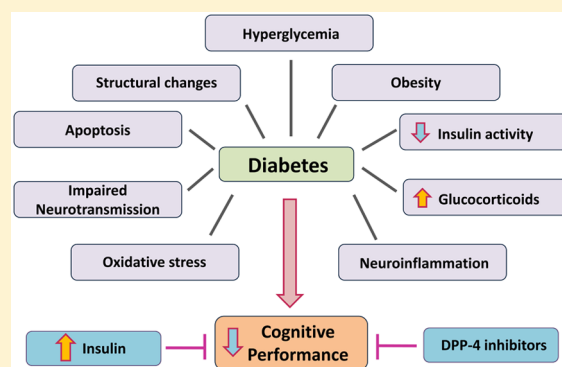
[§]Portuguese Diabetes Association (APDP), R. do Salitre 118-120, 1250-203 Lisboa, Portugal

^{||}CNC.IBILI Consortium, University of Coimbra, 3004-517 Coimbra, Portugal

[⊥]AIBILI, 3000-548 Coimbra, Portugal

ABSTRACT: Diabetes mellitus is the most common metabolic disease, and its prevalence is increasing. A growing body of evidence, both in animal models and epidemiological studies, has demonstrated that metabolic diseases like obesity, insulin resistance, and diabetes are associated with alterations in the central nervous system (CNS), being linked with development of cognitive and memory impairments and presenting a higher risk for dementia and Alzheimer's disease. The rising prevalence of diabetes together with its increasing earlier onset suggests that diabetes-related cognitive dysfunction will increase in the near future, causing substantial socioeconomic impact. Decreased insulin secretion or action, dysregulation of glucose homeostasis, impairment in the hypothalamic–pituitary–adrenal axis, obesity, hyperleptinemia, and inflammation may act independently or synergistically to disrupt neuronal homeostasis and cause diabetes-associated cognitive decline. However, the crosstalk between those factors and the mechanisms underlying the diabetes-related CNS complications is still elusive. During the past few years, different strategies (neuroprotective and antioxidant drugs) have emerged as promising therapies for this complication, which still remains to be preventable or treatable. This Review summarizes fundamental past and ongoing research on diabetes-associated cognitive decline, highlighting potential contributors, mechanistic mediators, and new pharmacological approaches to prevent and/or delay this complication.

KEYWORDS: Diabetes, hyperglycemia, insulinemia, obesity, brain, cognition, synapse



Diabetes mellitus, the most common metabolic disorder worldwide, is increasing dramatically. It is estimated that 387 million people live with diabetes, of which nearly 50% are unaware they have the disease mostly due to the fact that diabetes can remain asymptomatic or be misdiagnosed for several years (IDF diabetes Atlas 2014).

The underlying determinants of diabetes are the same all over the world. Economic development is associated with increasingly “obesogenic environments”, characterized by increasing access to energy-rich diets and decreased physical activity.

It is firmly established that diabetes causes micro- and macrovascular diseases.¹ Nevertheless, increasing evidence has also demonstrated that both type 1 and type 2 diabetes, in different degrees, result in complications in the central nervous system (CNS), with many brain structures being sensitive and responsive to changes in glucose homeostasis. Diabetes-associated cognitive decline (also known as diabetic encephalopathy) results from a complex interplay between direct and indirect metabolic consequences of long-term hyperglycemia,

insulin deficiency, and additional components such as genetic and environmental factors.²

Diabetes-associated cognitive decline has been linked to learning and memory deficits, which in turn increase the risk for dementia, Alzheimer's disease, and affective disorders.^{3,4} In this context, recent studies have shown that elevated body mass index, obesity, and insulin resistance are correlated with increased risk of dementia and cognitive impairment.^{5,6}

The mechanisms by which cognitive abilities are impaired in diabetes have not been clearly identified. Nevertheless, it has been shown that altered neurogenesis, electrophysiological deficits, oxidative stress injury, neuroinflammation, and neuronal apoptosis can induce structural changes and be involved in brain dysfunction during the course of diabetes.^{2,7}

This Review will give particular emphasis on the main factors underpinning type 1 and type 2 diabetic-related cognitive

Received: September 9, 2015

Accepted: December 15, 2015

Published: December 15, 2015

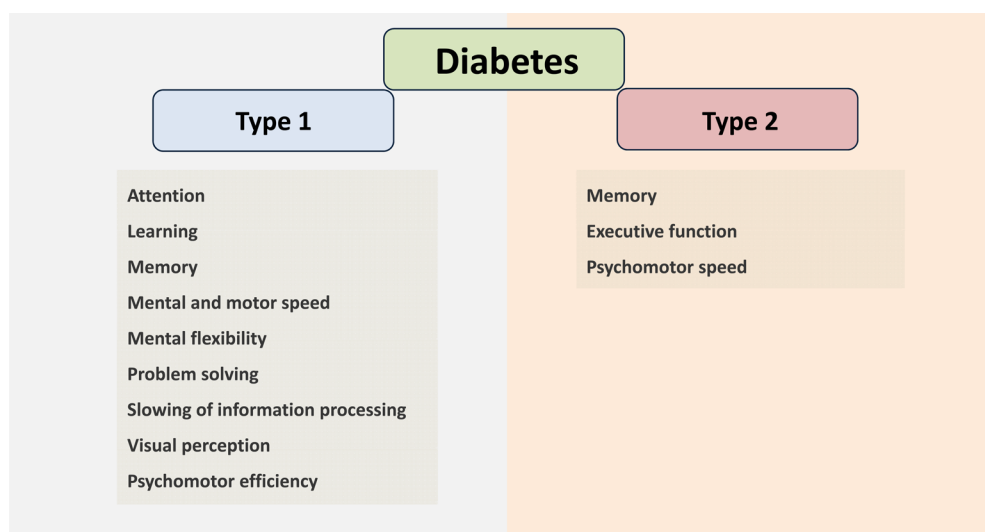


Figure 1. Diabetes-associated cognitive impairments described in type 1 and type 2 diabetic patients. Principal cognitive domains that have been found to be negatively affected in type 1 and type 2 diabetic patients.

deficits, highlighting recent advances on the treatment of diabetes-associated cognitive decline.

■ DIABETES AND COGNITIVE FUNCTION

Diabetes-associated cognitive decline has been associated with structural alterations in the brain, including brain atrophy, and electrophysiological deficits, that culminate in decreased memory and cognitive performance. The hippocampus, the brain region involved in memory formation and associative learning, is particularly affected by this pathology due to its sensitivity to changes in glucose and insulin homeostasis. The relative degree of cognitive dysfunction and the way cognitive abnormalities are manifested differs in both types of diabetes (Figure 1).^{4,7–10}

Type 1 diabetic patients have impairments in learning and memory, problem solving, and mental and motor speed. Type 1 diabetic children show declarative memory deficits,¹¹ whereas young adults have mild central brain atrophy, associated with changes in intellectual performance, mainly on information processing speed, mental flexibility, and psychomotor functioning.^{12,13} Moreover, 6 years after the onset of disease, children with type 1 diabetes have poorer performance on measures of intelligence, attention, processing speed, long-term memory, and executive skills comparing with control subjects.¹⁴

In animal models of type 1 diabetes, impaired performance in complex tasks, such as in Morris water maze or spatial-object learning task, has also been detected,^{15–18} with the development of behavioral deficits being dependent on diabetes duration.^{8,19} Intensive treatment with insulin¹⁶ or lowering corticosteroid levels^{17,20} prevents most of these deficits.

Patients with type 2 diabetes present moderate impairments in tasks involving verbal memory or complex information processing, increase in memory deficits, reduced frontal lobe/executive function, and reduction in psychomotor speed (Figure 1).^{14,21}

In animal models of insulin resistance, in which rats are submitted to a high fat or high sugar diets, learning impairments are detected, as assessed by hippocampus-dependent water maze tests. Also, insulin ability to induce long-term depression (LTD) is reduced in CA1 hippocampal subregion, which correlates with peripheral insulin resistance.²²

Moreover, rats fed with high-fat diet during 3 months present cognitive deficits,²³ while rats fed during 8 months exhibit impaired spatial learning ability, and reduced long-term potentiation (LTP) at Schaffer collateral - CA1 synapses.²⁴ In Zucker rats, an animal model of genetic obesity and insulin resistance, impaired spatial memory was also detected.²⁵

The differential effects induced by type 1 and type 2 diabetes on cognitive processes raise questions regarding the underlying causes and the progression of diabetes-associated cognitive decline.

■ MAIN CONTRIBUTORS TO DIABETES-ASSOCIATED COGNITIVE DECLINE

Hyperglycemia. Hyperglycemia is generally considered the major cause for diabetic complications, triggering several metabolic and molecular alterations that lead to progressive neuronal dysfunction.²⁶ Although the brain preferentially uses glucose as the main metabolic fuel source, under poor controlled diabetes (prolonged hyperglycemia), glucose transport is disturbed, which can lead to a dysregulation of brain metabolism and also to a dysregulation of osmolarity.^{26–28}

Higher plasma glucose levels (yet within the normal range) were associated with greater atrophy of the hippocampus and amygdala,²⁹ and accordingly, a better control of blood glucose levels improves cognitive performance in type 2 diabetic patients.²¹

Elevated glucose levels trigger various processes and activate several biochemical pathways that ultimately induce cell dysfunction and eventually cell death, slowly conducting to progressive functional and structural abnormalities in the brain.⁷ Dysfunctional glucose regulation elicits neuronal synaptic reorganization^{30,31} and increases proliferation of astrocytes³² in the hippocampus of streptozotocin (STZ)-induced diabetic animals. Synaptic reorganization includes depletion and dispersion of synaptic vesicles³¹ and changes in the content of proteins involved in exocytosis^{33,34} at presynaptic level, whereas an increase in postsynaptic density-95 (PSD-95) protein expression and changes in its distribution are described³⁰ at postsynaptic side. Nevertheless, hyperglycemia does not explain all the differences regarding neurological complications in type 1 and type 2 diabetes. In

fact, only a partial benefit is achieved when normal glycemia is maintained.³⁵ Nowadays, it is accepted that several factors (besides hyperglycemia) can act in additive or synergistic ways to impair neuronal homeostasis and increase neuronal vulnerability, thus contributing to cognitive decline. However, the interplay between those factors and the development of CNS complications remain to be clarified. The complex pathophysiological features of diabetes (Figure 2) may include

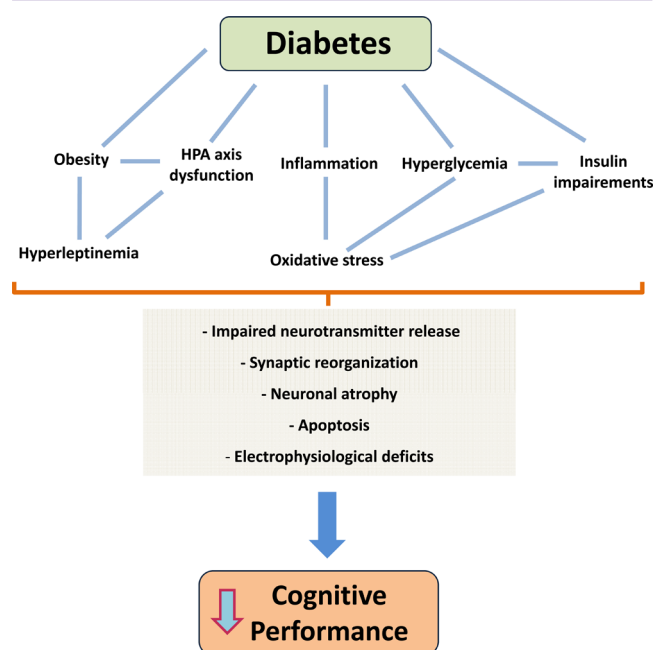


Figure 2. Main factors contributing to diabetes-induced brain dysfunction. The complex pathophysiological features of diabetes include decrease in insulin activity, impaired glucose homeostasis, and dysregulation of the hypothalamic–pituitary–adrenal axis function, among others. Changes observed in the brain under diabetic conditions include neuronal/dendritic atrophy, changes in synapse formation, impaired neurotransmitter release, and electrophysiological deficits.

not only a dysregulation of glucose homeostasis, but also decreased insulin secretion or action, impairment in the hypothalamic–pituitary–adrenal (HPA) axis, obesity, hyperleptinemia, oxidative stress, and inflammation.^{2,7}

Insulin Signaling. Hyperinsulinemia and insulin resistance occurs in the prediabetes and in early stages of type 2 diabetes, whereas chronic hypoinsulinemia occurs in type 1 and in later stages of type 2 diabetes. Insulin receptors are expressed throughout the brain suggesting an important role of insulin in the physiology of the CNS.

Insulin promotes proper synaptic metabolism, protein synthesis, neuronal survival, and the establishment of LTP (Figure 3A). Particularly in the hippocampus, insulin is a regulator of energy and glucose homeostasis,³⁶ hippocampal neuronal plasticity and cognitive functions (potentiating LTD and LTP),^{37–41} as well as neurogenesis.^{42–44} Insulin and the closely related proteins insulin-like growth factor (IGF)-1 and IGF-2 mediate their biological effects through the activation of two highly related tyrosine kinase receptors: the insulin receptor and the IGF-1 receptor. Both insulin and IGF-1 receptors are expressed throughout the brain, and use similar intracellular signaling machinery.

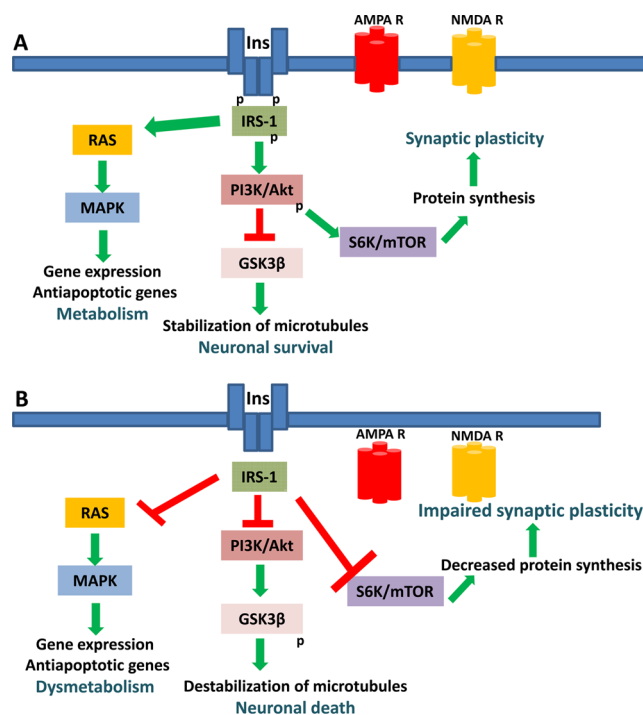


Figure 3. Neuronal insulin signaling. (A) Activation of insulin receptor leads to different signaling cascades with different physiological effects. Activation of MAPK signaling is responsible for the regulation of antiapoptotic gene expression and mitochondrial metabolism. Activation of Akt and consequently the inhibition of GSK-3 β leads to a microtubule stabilization and neuronal survival. Akt phosphorylation also activates mTOR pathway that is responsible for protein synthesis and the increase in synaptic plasticity. (B) Dysfunctional insulin signaling leads to dysmetabolism, impaired synaptic plasticity, destabilization of microtubules, and changes in the expression of genes involved in the apoptotic pathway.

Insulin receptor is a tyrosine kinase that activates downstream targets by phosphorylation of insulin receptor substrate (IRS). Phosphorylation of IRS leads to activation of phosphatidylinositol-3 kinase (PI3K) and mitogen-activated protein kinase (MAPK) pathways, which are involved in changes in hippocampal synaptic plasticity either by increasing protein synthesis or by stabilization of microtubules/axonal transport through GSK-3 β . Emerging data have shown that insulin may participate in the strengthening of synaptic plasticity through the activation of canonical PI3K. However, insulin might also activate other pathways such as S6K/mTOR that are involved in neuronal survival and nutrient sensing.^{41–43}

The multifactorial pathogenesis of diabetes-associated cognitive decline is not completely understood, but clearly shares features with brain aging and Alzheimer's disease.

In animal models of insulin resistance (high fat diet consumption), it has been described an impairment of insulin receptor function.²² As a result, changes in cytoskeleton components occur, leading to synaptic injury and neuronal loss; synaptic transmission is reduced and an attenuation of insulin-induced LTD is detected⁴⁵ (Figure 3B). Deficits in insulin signaling increase tau phosphorylation,^{46–48} a pathogenic feature detected in hippocampus of STZ diabetic mice and fatty Zucker rats, as well as in animal models of Alzheimer's disease.^{49,50} In addition, insulin treatment is able to improve cognitive performance,⁵¹ both in Alzheimer's disease⁵² and in nondemented individuals.⁵³

Alzheimer's disease is associated with the accumulation of neurofibrillary tangles and amyloid fibers, which leads to neuronal loss. Islet amyloid polypeptide (IAPP), which is cosecreted with insulin by pancreatic beta cells, spontaneously forms amyloid aggregates in a similar way of amyloid beta peptide. Degeneration of pancreatic islets, observed in type 2 diabetes, is associated with the formation of neurofibrillary tangles. These similarities in the accumulation of neurofibrillary tangles implicate a close biological relationship of protein aggregation between type 2 diabetes and Alzheimer's disease.⁵⁴

Several pieces of evidence indicate that insulin regulates the metabolism of amyloid beta and tau proteins. In the CNS, hypoinsulinemia can lower the levels of insulin-degrading enzyme (IDE), thereby impairing amyloid beta clearance. Also, chronic hyperinsulinemia in the peripheral circulation, along with decreased uptake of insulin into the brain, can lead to dysregulation of amyloid beta and inflammation.⁵⁵

Growing evidence also indicates that insulin regulates the metabolism of amyloid beta and tau proteins.⁵⁶ The levels of IDE are unchanged in the brain of diabetic animals,⁵⁷ although the increase in nitrosative stress can inhibit IDE.⁵⁸ IDE is an enzyme involved in insulin and amyloid beta protein degradation (the main component of amyloid plaques in the Alzheimer's brain). Inhibition of IDE promotes an accumulation of amyloid beta protein and leads to increased insulin levels and consequently insulin resistance.⁵⁸

Glucocorticoids. Although hyperglycemia and decreased insulin secretion or insulin activity are certainly central factors contributing to the complex pathophysiological characteristics of diabetes-associated cognitive decline, alterations in the HPA axis have also been shown to contribute for neuronal complications associated with diabetes. Elevated basal levels of glucocorticoids have been reported in patients with diabetes,^{59,60} which may influence proper CNS function and contribute for the development of cognitive impairments.^{61,62} A chronic increase in plasma glucocorticoids levels can aggravate symptoms induced by high blood glucose levels and enhance insulin resistance.⁶³ It was demonstrated that chronic elevation of corticosteroid levels induces changes in transcription of genes encoding corticotropin-releasing hormone receptors, suppresses dendritic arborization, and inhibits neurogenesis in the hippocampus.^{64,65}

In diabetic rodents, high levels of corticosteroids contribute to the impairment in synaptic plasticity and neurogenesis,^{17,41} induce depletion and clustering of synaptic vesicles in mossy fiber terminals,³¹ and increase the expression and the redistribution of synaptophysin in the hippocampus.³⁰ Maintaining physiological corticosterone levels and replacing insulin in diabetic rats prevents plasticity changes in the hippocampus.^{16,17}

Obesity and Hyperleptinemia. Both obesity and saturated fats are described to have damaging effects in the brain, being negatively correlated with cognitive performance. Obesity may contribute to dementia and other cognitive impairments by increasing the incidence of vascular pathologies,^{6,61,66} brain atrophy,⁶ and neuronal injury.⁶⁷

Excessive intake of certain macronutrients, such as simple carbohydrates and fatty acids, may lead to metabolic dysfunction, resulting also in alterations in structural plasticity and neurogenesis in some brain regions. In fact, the relation between obesity and central neuropathology has been demonstrated by evidence that weight reduction improves cognition.⁶⁸

In animals under high fat diet, a significant modification of neural and endocrine factors linked to energy and glucose homeostasis is observed. This, changes insulin action in the brain, even before the appearance of metabolic disease symptoms, therefore influencing the cognitive domain.⁶⁹

An additional consideration in obesity phenotypes is the potential for the increase in plasma levels of adipocyte derived hormones, such as leptin, that may have harmful consequences to the brain.⁵ Leptin receptors are abundantly expressed throughout the hippocampus, supporting a role for leptin in the facilitation of hippocampal synaptic plasticity under physiological conditions.⁷⁰ When directly administered in the CA1 hippocampal region, leptin improves memory processing and retention.⁷¹ Treatment of acute hippocampal slices with leptin results in the conversion of short-term potentiation to LTP by enhancing Ca²⁺ influx through NMDA receptors.⁷⁰ Leptin also increases neurogenesis in the dentate gyrus of adult mice and plays a critical role in hippocampal neuronal survival by activating the PI3K/Akt and JAK2/STAT3 signal transduction pathways.^{72,73} However, an increase in leptin levels observed under obesity induces long form receptor resistance thus impairing the signaling pathways and changing the normal synaptic plasticity.⁷⁴

Genetic mutations that disrupt leptin signaling such as in the db/db mouse and the Zucker fa/fa rat are associated with a reduction in LTP and impaired performance of hippocampal-dependent tasks,⁷⁵ suggesting that impairments in leptin signaling and leptin resistance contribute to deficits in hippocampal synaptic plasticity.

Oxidative Stress. Oxidative stress is defined as an increase in the steady-state levels of reactive oxygen species (ROS). Increased oxidative stress in diabetes is thought to promote the development of several diabetic complications. Diabetes may cause ROS production through glucose auto-oxidation, increased flux through the polyol pathway, and increases in protein glycation.²⁶ ROS may also activate aldose reductase and protein kinase C (PKC) and increase advanced glycation end-products (AGEs) and diacylglycerol formation.

In the brain of diabetic animals, there is an increase in oxidative and nitrosative stress, which can contribute to morphological abnormalities, neuronal damage, and cognitive and behavioral deficits.⁷⁶ Increased oxidative stress might result, not only from the increased generation of free radicals, but also from an impairment of the antioxidant defense system, responsible for scavenging free radicals and maintaining redox homeostasis. It was reported that together with the increase in ROS production, there is a decrease in the activity of antioxidant enzymes, namely superoxide dismutase and catalase, in the brain of diabetic rats.⁷⁷

Oxidative stress is linked to mitochondrial dysfunction that results in energy deficits and consequently neuronal damage and neurodegeneration.⁷⁸ Increasing evidence supports the idea that mitochondrial function declines with aging and with age related diseases; some authors consider brain mitochondrial dysfunction as the link between Alzheimer's disease and diabetes. Brain mitochondria isolated from STZ-induced diabetic rats have a lower content of CoQ9, which is a marker of antioxidant defense.⁷⁹ In a rat model of type 2 diabetes (Goto-Kakizaki), impairments in respiratory chain and oxidative phosphorylation in isolated brain mitochondria were reported.⁸⁰ Mitochondrial dysfunction in diabetes can be prevented/reduced by using antioxidants and insulin therapy.⁷⁹

Neuroinflammation. The increase in the expression of proinflammatory cytokines in the brain under diabetic conditions indicates that the innate immune system, and particularly microglial cells, are activated^{81,82} and play an important role in neuronal damage in diabetic patients and animals.^{83,84} The levels of proinflammatory cytokines are increased in the brain of patients and animal models with neuropathological disorders associated with cognitive function impairments, such as Alzheimer's disease and depression.^{85,86} In human postmortem hippocampus, it was detected that the activation of microglia in diabetic patients is similar to what is observed in Alzheimer's patients.⁸⁷

In the hippocampus of diabetic animals, the expression of tumor necrosis factor (TNF), interleukin-1 β (IL-1 β), IL-2, and IL-6 is increased.⁸¹ The increase in the expression of proinflammatory mediators in diabetic animals is associated with the increase of nitric oxide (NO) levels, simultaneously with the upregulation of the inducible nitric oxide synthase (iNOS) isoform, receptor for AGEs and NF- κ B.⁸² High fat diet fed mice present increased TNF levels and microglia/macrophage activation in the brain and adipose tissue, supporting the hypothesis that high fat diet dependent obesity results in concomitant proinflammatory changes in brain.⁸⁸ In type 2 diabetes, it was demonstrated that diabetic and obese mice (db/db) display impaired spatial-recognition memory, which is associated with increased levels of proinflammatory cytokines (IL-1 β , TNF, and IL-6) and reduced expression of BDNF in the hippocampus, thus suggesting an interaction between inflammation and memory impairment.⁸⁹

Moreover, it was found that after 2 weeks of diabetes there is an activation of microglia and enhanced GFAP expression in the hippocampus of STZ-induced diabetic rats.⁹⁰ Additionally, after 6 weeks of diabetes, increased caspase-3 activity induces profound cell death. This cell damage mediated by caspase 3 was concomitant with activation of astrocytes and microglia. The authors therefore suggested that these events are linked which further underpin the progression and severity of brain disorders, resulting in cognitive and behavioral impairments.⁹⁰

Inflammation, memory, and learning deficits are prevented in animals treated with antiinflammatory⁸³ and antioxidant⁸² compounds, and also in animals receiving C-peptide (peptide that connects A-chain of insulin to B-chain in the proinsulin molecule);⁸¹ however, the detailed molecular mechanism by which C-peptide has beneficial effects in the brain is still unclear. Recently, it was demonstrated that an antidepressant (Risperidone, phosphodiesterase-4 inhibitor) was able to decrease TNF levels and increase IL-10 levels, also preventing cognitive impairment in diabetic animals.⁹¹

A correlation between hyperglycemia, impaired insulin signaling, oxidative stress, and inflammation has been described. In diabetic encephalopathy, the physiological redox equilibrium is impaired leading to oxidative stress. This, in turn, is involved in multiple cellular pathways such as activation of transcription factors, AGE/RAGE, polyol, and PKC pathways, that are related to glucotoxicity and/or insulin signaling impairment. The ultimate consequence of such oxidative stress mechanisms related to diabetes is neuronal apoptosis and brain inflammation, which in turn are engaged with neurodegenerative events.²⁶

Another link between oxidative stress and insulin arises from impaired insulin signaling activity acting unfavorably on the expression and translocation of NF- κ B, which is also a modulator of ROS production. Activation of NF- κ B also

occurs in the presence of high glucose.⁹² NF- κ B regulates the expression of TNF and interleukins, playing a major role in the initiation of the inflammatory cascade. The upregulation of TNF has an inhibitory effect on insulin and IGF-1 signaling, thereby providing a self-perpetuating loop. NF- κ B signaling is also a modulator of apoptosis and ROS production and has been also associated with cognitive impairments.^{82,93}

■ NEUROSTRUCTURAL, NEUROCHEMICAL, AND ELECTROPHYSIOLOGICAL ALTERATIONS

Although several brain areas contribute to the cognitive deficits associated with diabetes, hippocampal-dependent tasks seem to be particularly sensitive to this disease.^{2,4,29,61,66,67} The multifactorial pathogenesis of diabetes-associated cognitive decline shares features with brain aging and Alzheimer's disease, such as alterations at neurochemical, structural, electrophysiological, and neurobehavioral levels. Altered regulation of the same intracellular signaling pathways was detected both in type 2 diabetes and in Alzheimer's disease. Based on these similarities, Alzheimer's disease is often referred as type 3 diabetes.

Structural Alterations and Neuronal Loss. The cognitive impairments in diabetic encephalopathy have been associated with structural alterations⁹⁴ and brain atrophy.^{13,95} Cortical, subcortical, and hippocampal atrophy (particularly in the dentate gyrus) has been detected in type 2 diabetic patients by brain magnetic resonance imaging.^{95–97}

In the STZ animal model of diabetes (type 1), swelling of neurons, glia, oligodendrocytes, synaptic boutons, and mitochondria has been reported in several brain regions.⁹⁴ Also, in the STZ animal model, diabetes reduces the density and length of neuronal dendrites^{98,99} and induces remodeling of dendrites which may be associated with the shrinkage of hippocampus.¹⁰⁰ In obese Zucker rats, a significant increase in the number of glial fibrillary acidic protein (GFAP) immunoreactive astrocytes was detected in the frontal and parietal cortex and also in the CA1 and CA3 subfields and dentate gyrus of the hippocampus, compared with the lean Zucker rats.¹⁰¹ Alterations of neurogenesis have also been reported in STZ-induced diabetic animals,^{102–105} and also in an animal model of type 2 diabetes (Goto-Kakizaki rats).¹⁰⁶

Moreover, apoptosis has also been implicated in diabetic complications. In vitro studies have shown that high glucose induces cell death in neuronal cells,^{107–109} and it was reported to occur in the hippocampus of diabetic animal models.^{110,111,103,112} In type 1 diabetic BioBreeding/Worcester (BB/Wor) rats, apoptosis was detected in CA1 and CA2 hippocampal neurons, which was accompanied by impairments in Morris water maze performance.^{110–112} Treatment with C-peptide was able to protect against hippocampal neuronal apoptosis, suggesting that cell death was in part caused by impaired insulin/C-peptide action/levels in BB/Wor rats.^{111,112}

Apoptosis was also detected in a type 2 diabetes animal model, BioBreedingZucker diabetic rat/Worcester (BBZDR/Wor), but in a lesser extent than in type 1 model, being more severe in CA1 pyramidal cells.¹¹² An increase in caspase-3 activity and Bax expression, and a significant increase in the ratio of Bax/Bcl-2 and Bax/Bcl-xL (used as index of apoptotic cell death) in the hippocampus of diabetic animals were also reported.¹¹³

Changes in Synaptic Transmission and Plasticity. In hippocampus of diabetic animal models, changes at presynaptic sites have been reported, including depletion and dispersion of

synaptic vesicles³¹ and changes in the content of proteins involved in exocytosis.^{33,34} In the postsynaptic terminal, it was detected an increase in PSD-95 protein expression, which may contribute to changes in the content and functional properties of glutamate receptors in the hippocampus of STZ diabetic rats.³⁰ PSD-95 controls the number of AMPA receptors at the synapses and participates in receptor delivery/stabilization during synaptic. Moreover, stronger synapses with increased PSD-95 levels showed enhanced LTD.¹¹⁴

Diabetes affects the synthesis and release of key neurotransmitters,^{115–119} which may underlie cognitive deficits observed in diabetic animals. It was reported in type 1 animal models of diabetes, increased levels of dopamine in different brain areas,¹²⁰ decreased basal levels of serotonin and dopamine in the hippocampus, and also decreased basal levels of glutamate in dentate gyrus.¹²¹ The basal levels of GABA in the hippocampus are not altered in STZ-induced diabetic animals.^{119,122} However, under chronic hyperglycemia, extracellular brain levels of GABA and glutamate are decreased.¹¹⁵ In addition, GABA receptors are downregulated in STZ-induced diabetic animals,¹²³ which may contribute to changes in inhibitory function and disruption of memory. Imbalances between excitatory and inhibitory neurotransmission are triggering factors for neurodegeneration and consequently may contribute to changes in cognitive processes.

Acetylcholine esterase has a fundamental role in cognitive processes, and any alterations in its activity, as well as in the neurotransmitter acetylcholine, are associated with cognitive deficits observed in patients and animal models of diabetes.¹²⁴ In the hippocampus of type 1 STZ diabetic animals, acetylcholine esterase mRNA expression increases, whereas choline acetyltransferase mRNA expression decreases, suggesting a reduction in cholinergic transmission in hippocampal neurons.¹²³

ATP signaling is also compromised in the hippocampus of STZ-induced diabetic rats. There is a decrease in ATP release and a downregulation of synaptic P2 receptors.¹²⁵ Extracellular ATP plays a role in synaptic efficiency processes by the activation of P2 receptors at the presynaptic sites, to control the release of glutamate,¹²⁶ and at the postsynaptic cell facilitating the activation of ionotropic glutamate receptors.¹²⁷

Alterations in Ca²⁺ homeostasis and Ca²⁺-dependent forms of synaptic plasticity in the hippocampus of diabetic animals have been associated with alterations in neurotransmitter release and learning impairments. Increasing evidence reveals that both type 1 and type 2 diabetes impairs Ca²⁺ homeostasis;^{128–130} small changes in free intracellular Ca²⁺ sustained over a period of time results in cellular damage.¹³⁰ Cellular Ca²⁺ homeostasis is required for proper neuronal function and mitochondria has a major role in regulating intracellular Ca²⁺. In fact, excessive Ca²⁺ uptake into mitochondria has been shown to increase ROS production, inhibit ATP synthesis, and induce mitochondrial permeability transition (MPT). MPT results in mitochondrial swelling and can lead to the release of proapoptotic proteins. Mitochondrial dysfunction and the resulting energy deficit trigger the onset of neuronal degeneration and death.⁷⁸

Furthermore, Ca²⁺ currents and the expression of voltage-sensitive Ca²⁺ channels^{131–133} are increased in dorsal root ganglion neurons from BB/W and STZ-induced diabetic rats. In retinal neurons exposed to high glucose, the increase in the intracellular Ca²⁺ concentration evoked by cell depolarization is

enhanced, and the recovery to basal Ca²⁺ levels is delayed comparing with control retinal neurons.¹³⁴

Slow afterhyperpolarization (Ca²⁺-dependent feature of synaptic transmission occurring following a sequence of action potentials) is increased in the hippocampus of diabetic rats.¹²⁸ In an established animal model of brain aging (F344 rats), high-fat diet exacerbates cognitive decline and the hippocampal Ca²⁺-dependent afterhyperpolarization (a marker of age-dependent Ca²⁺ dysregulation).¹³⁵

Diabetes-induced alterations in synaptic neurotransmission at pre- and postsynaptic level may lead to changes in synaptic plasticity (Figure 4). Impairments in hippocampal synaptic

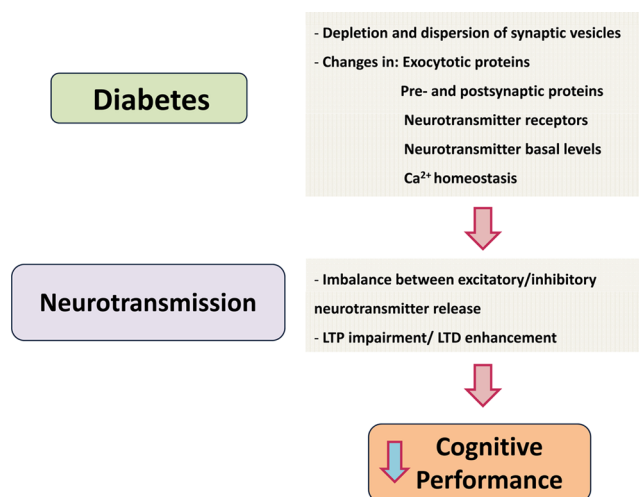


Figure 4. Neurotransmission impairment under diabetes correlates with decreased cognitive performance. Diabetes affects the synapse both at pre- and postsynaptic level, which may underlie changes in synaptic plasticity and consequently cognitive deficits that have been detected in diabetic patients and animal models.

plasticity and cognition have been demonstrated in several animal models of diabetes, such as impaired LTP and enhanced LTD,^{16,17,136,137} although in the STZ model, LTP does not occur in more than 50% of animals. Moreover, when LTP is induced in diabetic animals, the magnitude of the excitatory postsynaptic potential (EPSP) slope is slower and is not sustained comparing with control animals.¹⁶

LTP induction is regulated by postsynaptic NMDA receptors. NMDA receptors containing the NR2B subunit are decreased in the hippocampus of STZ-induced diabetic animals, causing a decrease in EPSPs.^{138,139} Besides the reduction of the NR2B subunit, its phosphorylation by CaMKII also decreases in postsynaptic densities.¹³⁸ The affinity of glutamate for AMPA receptors, but not for NMDA receptors, is decreased in the hippocampus of STZ-induced diabetic rats.^{140,141} In STZ-diabetic animals, the transient and the persistent sodium currents are decreased and the potassium currents are increased in hippocampal CA1 neurons, contributing to the decrease in membrane excitability. Moreover, the resting membrane potential of diabetic neurons was more positive than the control neurons.¹⁰

Taken together, changes reported in neurotransmitter release suggest the existence of an imbalance between excitation and inhibition, thus contributing to neuronal dysfunction and ultimately to memory impairments detected in diabetic animals and humans.

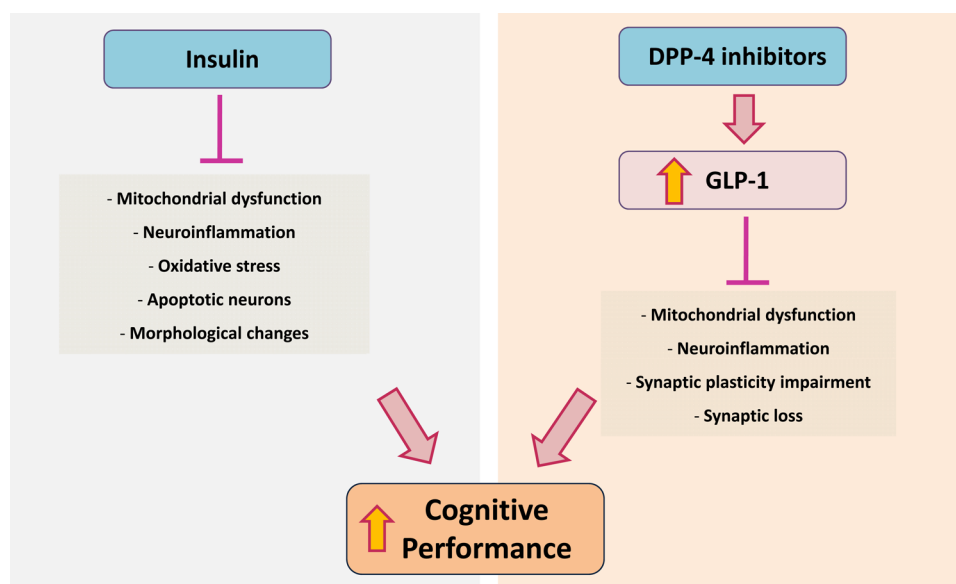


Figure 5. Pharmacological therapeutic strategies in diabetic encephalopathy. Insulin therapy and increasing GLP-1 levels using DPP-4 inhibitors can improve cognitive performance under diabetes through several mechanisms, attenuating brain mitochondrial dysfunction, neuroinflammation, and synaptic/neuronal loss, among others.

■ PHARMACOLOGICAL THERAPEUTIC STRATEGIES

Unraveling the pathogenesis of diabetes-triggered dysfunction in the brain will open new avenues for devising possible innovative pharmacological approaches to treat diabetes-associated cognitive decline. During the past few years, different strategies have been considered for the prevention or treatment of this complication; however, diabetic encephalopathy still remains to be preventable or treatable. Given the underlying mechanisms of diabetes-associated cognitive decline, the neuroprotective strategies have been focused mainly on the reduction of oxidative stress and the prevention of apoptosis to avoid neuronal loss.

Antioxidant and Neuroprotective Agents. As oxidative stress is one of the factors that can induce hippocampal dysfunction and consequently cognitive dysfunction, several studies have reported the protective effect of antioxidants in hippocampus of animal models of diabetes and obesity. Isoflavones have a metabolic action on energy balance and, due to its antioxidant properties, protects cultured primary hippocampal neurons from oxidative stress induced by exposure to glutamate and beta-amyloid. Treatment of STZ-induced diabetic animals with melatonin and vitamin E significantly ameliorates learning and memory performance, and can reverse lipid peroxidation and glutathione levels toward control values.¹⁴²

Sesamol (3,4-methylenedioxyphenol) is a phenolic antioxidant and anti-inflammatory molecule that attenuates cognitive deficits and reduces acetylcholinesterase activity, oxidative stress, and inflammation in STZ-induced diabetic rats.⁸³ These authors also demonstrated that chronic treatment with curcumin, a well-established phenolic antioxidant and anti-inflammatory molecule, significantly attenuates cognitive deficits, oxidative stress, and inflammation in STZ-induced diabetic rats.¹⁴³

Moreover, N-acetylcysteine administration for 8 weeks significantly ameliorates oxidative stress, improves lipid composition, and restores membrane fluidity and the activity of membrane bound enzymes in STZ-induced diabetic rats.¹⁴⁴

Another molecule, etomidate, significantly decreases nitrite levels in the rat cortex, hippocampus, cerebellum, brain stem, and spinal cord, demonstrating that etomidate can have neuroprotective effects on the neuronal tissue against the diabetic-induced oxidative damage.¹⁴⁵

Insulin and C-Peptide. In the brain, insulin has neurotrophic and neuroprotective effects, facilitating synaptic plasticity in the hippocampus and enhancing memory function. Based on experimental and clinical studies that support important roles for insulin in several neuronal functions, particularly in plasticity events, insulin is currently being studied as a potential therapeutic strategy for cognitive deficits (Figure 5). Insulin is able to protect against oxidative stress and the reduction of mitochondrial oxidative phosphorylation efficiency induced by amyloid beta peptide in STZ rats.⁷⁹ Moreover, insulin treatment is able to prevent morphological changes in hippocampal CA3 subregion,¹⁴⁶ as well as the decrease in GFAP levels and the increase in glutamate uptake in diabetic animal models.¹⁴⁷ Moreover, replacement of insulin in type 1 diabetic rats protects against hippocampal damage and prevents cognitive deficits.¹⁶

Insulin and/or C-peptide prevent apoptotic neuronal cell death mediated by PI3-kinase stimulation. They also prevent the p38 activation and the disinhibition of NF- κ B translocation^{111,148} and the release of proinflammatory factors in the hippocampus of diabetic animals.⁸¹

Even though insulin therapy can be useful to prevent or slow the progression of diabetic complications, it is important to keep in mind that these hormone beneficial effects may be compromised by the increased risk of severe hypoglycemic episodes, which in turn are associated with increased neuronal and cognitive dysfunction. Alternatively, the use of insulin sensitizers may provide a reliable therapeutic alternative.

Dipeptidyl Peptidase-4 Inhibitors and GLP-1 Agonists. Glucagon-like peptide-1 (GLP-1) facilitates insulin release from the pancreas under hyperglycemic conditions. However, its short half-life is a limitation for its use as a therapeutic tool. Based on this, long-acting glucagon-like

peptide-1 receptor (GLP-1R) agonists and dipeptidyl peptidase-4 (DPP-4) inhibitors have been developed as treatments for type 2 diabetes. These drugs increase insulin secretion and decrease glucagon production in a glucose-dependent manner without promoting hypoglycemia. GLP-1 analogues and DPP-4 inhibitors have protective effects in several tissues, including the brain (Figure 5).

GLP-1 receptors are expressed throughout the brain and GLP-1 can cross the blood-brain barrier (BBB), suggesting that they are promising therapeutic targets in neurodegenerative diseases, such as diabetes-associated cognitive decline and Alzheimer's disease.¹⁴⁹ Agonists of GLP-1R have anti-inflammatory and neurogenic effects, and are able to reduce beta-amyloid plaques preventing loss of synapses and memory impairments.^{150–152}

Vildagliptin and nalogliptin (DPP-4 inhibitors) increase plasma and brain GLP-1 levels and restore insulin-induced long-term depression. They also restore neuronal insulin receptor phosphorylation, IRS-1 phosphorylation, and Akt/PKB-ser phosphorylation, attenuating the impaired cognitive function caused by high fat diet.^{153,154} Vildagliptin and sitagliptin completely restore brain mitochondrial function and improve learning and memory behaviors in rats with high fat diet induced insulin resistance.¹⁵⁵ Liraglutide (long-acting GLP-1 agonist) prevents or attenuates type 2 diabetes associated neuronal and cognitive deficits,¹⁵¹ possibly by a mechanism that promotes neurogenesis in dentate gyrus.¹⁵⁶ Liraglutide significantly affects brain neurotransmission and modulates synaptic plasticity enhancing LTP; it also protects against impairment in learning and memory in animal models of Alzheimer's disease and diabetes.¹⁵⁷ In a type 2 diabetic animal model, liraglutide normalizes brain metabolic homeostasis, decreases hippocampal lipid oxidation, improves brain mitochondrial regulation, and preserves synaptic plasticity (for a review, see ref 149). In line with these studies, two clinical trials are ongoing to analyze the effect of liraglutide as a neuroprotective drug in Alzheimer's disease.

CONCLUSIONS

Measurable manifestations of diabetic encephalopathy include impairment in cognitive functions and brain structural and electrophysiological changes. The precise mechanisms that underpin this pathology remain to be fully determined. Nonetheless, they are likely to involve impaired insulin signaling and increased inflammatory and oxidative stress pathways; these events occur early in the neurodegenerative process and share complex features with mechanisms of brain aging.

New treatment approaches for managing diabetes have shown some efficacy in reducing diabetes-induced neurodegeneration in animal studies, and promising drugs are being assessed to prevent or slow the progression of the disease. In addition, the management of diabetes can often involve a combination of treatments to control hyperglycemia and hyperinsulinemia, as well as treatments to improve peripheral insulin sensitivity. Advances in the understanding of the mechanisms underlying cognitive deficits pave the way to develop novel neuroprotective strategies aiming at preventing, delaying, and/or reversing the neurological consequences of diabetes.

AUTHOR INFORMATION

Corresponding Author

*Phone: + 351 21 880 30 17. Fax: + 351 21 880 3028. E-mail: joanamgaspar@gmail.com.

Author Contributions

J.M.G. designed, wrote and edited the manuscript; F.I.B. wrote and critically reviewed the manuscript; M.P.M. and A.F.A. contributed to critical review of the manuscript.

Funding

F.I.B. acknowledges the fellowship from Fundação para a Ciência e a Tecnologia, Portugal (SFRH/BPD/86830/2012).

Notes

The authors declare no competing financial interest.

ABBREVIATIONS

BDNF, brain derived neurotrophic factor; CA, *CornuAmmonis*; CNS, central nervous system; DPP-4, dipeptidylpeptidase 4; EPSP, excitatory postsynaptic potential; GFAP, Glial fibrillary acidic protein; GLP-1, glucagon-like peptide-1; HPA, hypothalamic-pituitary-adrenal; IDE, insulin degrading enzyme; IGF, insulin-like growth factor; IL, interleukin; LTD, long-term depression; LTP, long-term potentiation; NO, nitric oxide; ROS, reactive oxygen species; STZ, streptozotocin; TNF, tumor necrosis factor

REFERENCES

- (1) American Diabetes Association (2013) Diagnosis and classification of diabetes mellitus. *Diabetes Care* 36 (Suppl 1), S67–74.
- (2) Stranahan, A. M. (2015) Models and mechanisms for hippocampal dysfunction in obesity and diabetes. *Neuroscience* 309, 125.
- (3) Biessels, G. J., Kappelle, A. C., Bravenboer, B., Erkelens, D. W., and Gispen, W. H. (1994) Cerebral function in diabetes mellitus. *Diabetologia* 37, 643–650.
- (4) Yates, K. F., Sweat, V., Yau, P. L., Turchiano, M. M., and Convit, A. (2012) Impact of metabolic syndrome on cognition and brain: a selected review of the literature. *Arterioscler., Thromb., Vasc. Biol.* 32, 2060–2067.
- (5) Grillo, C. A., Piroli, G. G., Junor, L., Wilson, S. P., Mott, D. D., Wilson, M. A., and Reagan, L. P. (2011) Obesity/hyperleptinemic phenotype impairs structural and functional plasticity in the rat hippocampus. *Physiol. Behav.* 105, 138.
- (6) Raji, C. A., Ho, A. J., Parikshak, N. N., Becker, J. T., Lopez, O. L., Kuller, L. H., Hua, X., Leow, A. D., Toga, A. W., and Thompson, P. M. (2010) Brain structure and obesity. *Hum Brain Mapp.* 31, 353–364.
- (7) Wrighten, S. A., Piroli, G. G., Grillo, C. A., and Reagan, L. P. (2009) A look inside the diabetic brain: Contributors to diabetes-induced brain aging. *Biochim. Biophys. Acta, Mol. Basis Dis.* 1792, 444–453.
- (8) Gispen, W. H., and Biessels, G. J. (2000) Cognition and synaptic plasticity in diabetes mellitus. *Trends Neurosci.* 23, 542–549.
- (9) Gragnoli, C. (2012) Depression and type 2 diabetes: cortisol pathway implication and investigational needs. *J. Cell. Physiol.* 227, 2318–2322.
- (10) Heng, L. J., Yang, R. H., and Jia, D. (2011) Diabetes impairs learning performance through affecting membrane excitability of hippocampal pyramidal neurons. *Behav. Brain Res.* 224, 250–258.
- (11) Hershey, T., Lillie, R., Sadler, M., and White, N. H. (2004) A prospective study of severe hypoglycemia and long-term spatial memory in children with type 1 diabetes. *Pediatr. Diabetes* 5, 63–71.
- (12) Brands, A. M., Biessels, G. J., de Haan, E. H., Kappelle, L. J., and Kessels, R. P. (2005) The effects of type 1 diabetes on cognitive performance: a meta-analysis. *Diabetes Care* 28, 726–735.
- (13) Ferguson, S. C., Blane, A., Wardlaw, J., Frier, B. M., Perros, P., McCrimmon, R. J., and Deary, I. J. (2005) Influence of an early-onset

age of type 1 diabetes on cerebral structure and cognitive function. *Diabetes Care* 28, 1431–1437.

(14) Kodl, C. T., and Seaquist, E. R. (2008) Cognitive dysfunction and diabetes mellitus. *Endocr. Rev.* 29, 494–511.

(15) Biessels, G., and Gispen, W. H. (1996) The calcium hypothesis of brain aging and neurodegenerative disorders: significance in diabetic neuropathy. *Life Sci.* 59, 379–387.

(16) Biessels, G. J., Kamal, A., Urban, I. J., Spruijt, B. M., Erkelens, D. W., and Gispen, W. H. (1998) Water maze learning and hippocampal synaptic plasticity in streptozotocin-diabetic rats: effects of insulin treatment. *Brain Res.* 800, 125–135.

(17) Stranahan, A. M., Arumugam, T. V., Cutler, R. G., Lee, K., Egan, J. M., and Mattson, M. P. (2008) Diabetes impairs hippocampal function through glucocorticoid-mediated effects on new and mature neurons. *Nat. Neurosci.* 11, 309–317.

(18) Popovic, M., Biessels, G. J., Isaacson, R. L., and Gispen, W. H. (2001) Learning and memory in streptozotocin-induced diabetic rats in a novel spatial/object discrimination task. *Behav. Brain Res.* 122, 201–207.

(19) Sima, A. A., Kamiya, H., and Li, Z. G. (2004) Insulin, C-peptide, hyperglycemia, and central nervous system complications in diabetes. *Eur. J. Pharmacol.* 490, 187–197.

(20) Stranahan, A. M., Lee, K., Pistell, P. J., Nelson, C. M., Readal, N., Miller, M. G., Spangler, E. L., Ingram, D. K., and Mattson, M. P. (2008) Accelerated cognitive aging in diabetic rats is prevented by lowering corticosterone levels. *Neurobiol. Learn. Mem.* 90, 479–483.

(21) Ryan, C. M., Freed, M. I., Rood, J. A., Cobitz, A. R., Waterhouse, B. R., and Strachan, M. W. (2006) Improving metabolic control leads to better working memory in adults with type 2 diabetes. *Diabetes Care* 29, 345–351.

(22) Pratchayasakul, W., Kerdphoo, S., Petsophonakul, P., Pongchaidecha, A., Chattipakorn, N., and Chattipakorn, S. C. (2011) Effects of high-fat diet on insulin resistance function in rat hippocampus and the level of neuronal corticosterone. *Life Sci.* 88, 619–627.

(23) Greenwood, C. E., and Winocur, G. (1996) Cognitive impairment in rats fed high-fat diets: a specific effect of saturated fatty-acid intake. *Behav. Neurosci.* 110, 451–459.

(24) Stranahan, A. M., Norman, E. D., Lee, K., Cutler, R. G., Telljohann, R. S., Egan, J. M., and Mattson, M. P. (2008) Diet-induced insulin resistance impairs hippocampal synaptic plasticity and cognition in middle-aged rats. *Hippocampus* 18, 1085.

(25) Winocur, G., Greenwood, C. E., Piroli, G. G., Grillo, C. A., Reznikov, L. R., Reagan, L. P., and McEwen, B. S. (2005) Memory impairment in obese Zucker rats: an investigation of cognitive function in an animal model of insulin resistance and obesity. *Behav. Neurosci.* 119, 1389–1395.

(26) Tomlinson, D. R., and Gardiner, N. J. (2008) Glucose neurotoxicity. *Nat. Rev. Neurosci.* 9, 36–45.

(27) Seaquist, E. R., Tkac, I., Damberg, G., Thomas, W., and Gruetter, R. (2005) Brain glucose concentrations in poorly controlled diabetes mellitus as measured by high-field magnetic resonance spectroscopy. *Metab., Clin. Exp.* 54, 1008–1013.

(28) Vannucci, S. J., Gibbs, E. M., and Simpson, I. A. (1997) Glucose utilization and glucose transporter proteins GLUT-1 and GLUT-3 in brains of diabetic (db/db) mice. *Am. J. Physiol.* 272, E267–274.

(29) Cherbuin, N., Sachdev, P., and Anstey, K. J. (2012) Higher normal fasting plasma glucose is associated with hippocampal atrophy: The PATH Study. *Neurology* 79, 1019–1026.

(30) Grillo, C. A., Piroli, G. G., Wood, G. E., Reznikov, L. R., McEwen, B. S., and Reagan, L. P. (2005) Immunocytochemical analysis of synaptic proteins provides new insights into diabetes-mediated plasticity in the rat hippocampus. *Neuroscience* 136, 477–486.

(31) Magarinos, A. M., and McEwen, B. S. (2000) Experimental diabetes in rats causes hippocampal dendritic and synaptic reorganization and increased glucocorticoid reactivity to stress. *Proc. Natl. Acad. Sci. U. S. A.* 97, 11056–11061.

(32) Saravia, F. E., Revsin, Y., Gonzalez Deniselle, M. C., Gonzalez, S. L., Roig, P., Lima, A., Homo-Delarche, F., and De Nicola, A. F. (2002)

Increased astrocyte reactivity in the hippocampus of murine models of type 1 diabetes: the nonobese diabetic (NOD) and streptozotocin-treated mice. *Brain Res.* 957, 345–353.

(33) Duarte, J. M., Carvalho, R. A., Cunha, R. A., and Gruetter, R. (2009) Caffeine consumption attenuates neurochemical modifications in the hippocampus of streptozotocin-induced diabetic rats. *J. Neurochem.* 111, 368–379.

(34) Gaspar, J. M., Baptista, F. I., Galvao, J., Castilho, A. F., Cunha, R. A., and Ambrosio, A. F. (2010) Diabetes differentially affects the content of exocytotic proteins in hippocampal and retinal nerve terminals. *Neuroscience* 169, 1589–1600.

(35) Sima, A. A., and Kamiya, H. (2008) Is C-peptide replacement the missing link for successful treatment of neurological complications in type 1 diabetes? *Curr. Drug Targets* 9, 37–46.

(36) Air, E. L., Benoit, S. C., Clegg, D. J., Seeley, R. J., and Woods, S. C. (2002) Insulin and leptin combine additively to reduce food intake and body weight in rats. *Endocrinology* 143, 2449–2452.

(37) Zhao, W. Q., and Alkon, D. L. (2001) Role of insulin and insulin receptor in learning and memory. *Mol. Cell. Endocrinol.* 177, 125–134.

(38) Man, H. Y., Lin, J. W., Ju, W. H., Ahmadian, G., Liu, L., Becker, L. E., Sheng, M., and Wang, Y. T. (2000) Regulation of AMPA receptor-mediated synaptic transmission by clathrin-dependent receptor internalization. *Neuron* 25, 649–662.

(39) Ahmadian, G., Ju, W., Liu, L., Wyszynski, M., Lee, S. H., Dunah, A. W., Taghibiglou, C., Wang, Y., Lu, J., Wong, T. P., Sheng, M., and Wang, Y. T. (2004) Tyrosine phosphorylation of GluR2 is required for insulin-stimulated AMPA receptor endocytosis and LTD. *EMBO J.* 23, 1040–1050.

(40) van der Heide, L. P., Kamal, A., Artola, A., Gispen, W. H., and Ramakers, G. M. (2005) Insulin modulates hippocampal activity-dependent synaptic plasticity in a N-methyl-D-aspartate receptor and phosphatidylinositol-3-kinase-dependent manner. *J. Neurochem.* 94, 1158–1166.

(41) Lee, C. C., Huang, C. C., and Hsu, K. S. (2011) Insulin promotes dendritic spine and synapse formation by the PI3K/Akt/mTOR and Rac1 signaling pathways. *Neuropharmacology* 61, 867–879.

(42) Llorens-Martin, M., Torres-Aleman, I., and Trejo, J. L. (2009) Mechanisms mediating brain plasticity: IGF1 and adult hippocampal neurogenesis. *Neuroscientist* 15, 134–148.

(43) Liu, W., Ye, P., O'Kusky, J. R., and D'Ercole, A. J. (2009) Type 1 insulin-like growth factor receptor signaling is essential for the development of the hippocampal formation and dentate gyrus. *J. Neurosci. Res.* 87, 2821–2832.

(44) Guo, J., Yu, C., Li, H., Liu, F., Feng, R., Wang, H., Meng, Y., Li, Z., Ju, G., and Wang, J. (2010) Impaired neural stem/progenitor cell proliferation in streptozotocin-induced and spontaneous diabetic mice. *Neurosci. Res.* 68, 329–336.

(45) Pipatpiboon, N., Pratchayasakul, W., Chattipakorn, N., and Chattipakorn, S. C. (2012) PPARgamma Agonist Improves Neuronal Insulin Receptor Function in Hippocampus and Brain Mitochondria Function in Rats with Insulin Resistance Induced by Long Term High-Fat Diets. *Endocrinology* 153, 329–338.

(46) Liu, Y., Liu, F., Grundke-Iqbal, I., Iqbal, K., and Gong, C. X. (2011) Deficient brain insulin signalling pathway in Alzheimer's disease and diabetes. *J. Pathol.* 225, 54–62.

(47) Iqbal, K., Liu, F., Gong, C. X., Alonso, A. d. C., and Grundke-Iqbal, I. (2009) Mechanisms of tau-induced neurodegeneration. *Acta Neuropathol.* 118, 53–69.

(48) Freude, S., Plum, L., Schnitker, J., Leiser, U., Udelhoven, M., Krone, W., Bruning, J. C., and Schubert, M. (2005) Peripheral hyperinsulinemia promotes tau phosphorylation in vivo. *Diabetes* 54, 3343–3348.

(49) Kim, B., Backus, C., Oh, S., Hayes, J. M., and Feldman, E. L. (2009) Increased tau phosphorylation and cleavage in mouse models of type 1 and type 2 diabetes. *Endocrinology* 150, S294–S301.

(50) Qu, Z., Jiao, Z., Sun, X., Zhao, Y., Ren, J., and Xu, G. (2011) Effects of streptozotocin-induced diabetes on tau phosphorylation in the rat brain. *Brain Res.* 1383, 300–306.

- (51) Francis, G. J., Martinez, J. A., Liu, W. Q., Xu, K., Ayer, A., Fine, J., Tuor, U. I., Glazner, G., Hanson, L. R., Frey, W. H., 2nd, and Toth, C. (2008) Intranasal insulin prevents cognitive decline, cerebral atrophy and white matter changes in murine type I diabetic encephalopathy. *Brain* 131, 3311–3334.
- (52) Reger, M. A., Watson, G. S., Green, P. S., Baker, L. D., Cholerton, B., Fishel, M. A., Plymate, S. R., Cherrier, M. M., Schellenberg, G. D., Frey, W. H., 2nd, and Craft, S. (2008) Intranasal insulin administration dose-dependently modulates verbal memory and plasma amyloid-beta in memory-impaired older adults. *J. Alzheimer's Dis.* 13, 323–331.
- (53) Benedict, C., Hallschmid, M., Schultes, B., Born, J., and Kern, W. (2007) Intranasal insulin to improve memory function in humans. *Neuroendocrinology* 86, 136–142.
- (54) Sridhar, G. R., Lakshmi, G., and Nagamani, G. (2015) Emerging links between type 2 diabetes and Alzheimer's disease. *World J. Diabetes* 6, 744–751.
- (55) Qiu, W. Q., and Folstein, M. F. (2006) Insulin, insulin-degrading enzyme and amyloid-beta peptide in Alzheimer's disease: review and hypothesis. *Neurobiol. Aging* 27, 190–198.
- (56) Craft, S., and Watson, G. S. (2004) Insulin and neurodegenerative disease: shared and specific mechanisms. *Lancet Neurol.* 3, 169–178.
- (57) Azam, M., Gupta, B. L., Gupta, G., Jain, S., and Baquer, N. Z. (1990) Rat brain insulin degrading enzyme in insulin and thyroid hormones imbalances. *Biochem Int.* 21, 321–329.
- (58) Kummer, M. P., Hulsmann, C., Hermes, M., Axt, D., and Heneka, M. T. (2012) Nitric oxide decreases the enzymatic activity of insulin degrading enzyme in APP/PS1 mice. *J. Neuroimmune Pharmacol.* 7, 165–172.
- (59) Reagan, L. P., Grillo, C. A., and Piroli, G. G. (2008) The As and Ds of stress: metabolic, morphological and behavioral consequences. *Eur. J. Pharmacol.* 585, 64–75.
- (60) Couch, R. M. (1992) Dissociation of cortisol and adrenal androgen secretion in poorly controlled insulin-dependent diabetes mellitus. *Eur. J. Endocrinol.* 127, 115–117.
- (61) Bruehl, H., Wolf, O. T., Sweat, V., Tirsi, A., Richardson, S., and Convit, A. (2009) Modifiers of cognitive function and brain structure in middle-aged and elderly individuals with type 2 diabetes mellitus. *Brain Res.* 1280, 186–194.
- (62) Beauquis, J., Homo-Delarche, F., Revsin, Y., De Nicola, A. F., and Saravia, F. (2008) Brain alterations in autoimmune and pharmacological models of diabetes mellitus: focus on hypothalamic-pituitary-adrenocortical axis disturbances. *Neuroimmunomodulation* 15, 61–67.
- (63) Yi, S. S., Hwang, I. K., Chun, M. S., Kim, Y. N., Kim, I. Y., Lee, I. S., Seong, J. K., and Yoon, Y. S. (2009) Glucocorticoid receptor changes associate with age in the paraventricular nucleus of type II diabetic rat model. *Neurochem. Res.* 34, 851–858.
- (64) Joels, M., Karst, H., Krugers, H. J., and Lucassen, P. J. (2007) Chronic stress: implications for neuronal morphology, function and neurogenesis. *Front. Neuroendocrinol.* 28, 72–96.
- (65) Joels, M., and Baram, T. Z. (2009) The neuro-symphony of stress. *Nat. Rev. Neurosci.* 10, 459–466.
- (66) Bruehl, H., Sweat, V., Tirsi, A., Shah, B., and Convit, A. (2011) Obese Adolescents with Type 2 Diabetes Mellitus Have Hippocampal and Frontal Lobe Volume Reductions. *Neurosci. Med.* 2, 34–42.
- (67) Mueller, K., Sacher, J., Arelin, K., Holiga, S., Kratzsch, J., Villringer, A., and Schroeter, M. L. (2012) Overweight and obesity are associated with neuronal injury in the human cerebellum and hippocampus in young adults: a combined MRI, serum marker and gene expression study. *Transl. Psychiatry* 2, e200.
- (68) Park, H. R., and Lee, J. (2011) Neurogenic contributions made by dietary regulation to hippocampal neurogenesis. *Ann. N. Y. Acad. Sci.* 1229, 23–28.
- (69) Banas, S. M., Rouch, C., Kassis, N., Markaki, E. M., and Gerozissis, K. (2009) A dietary fat excess alters metabolic and neuroendocrine responses before the onset of metabolic diseases. *Cell. Mol. Neurobiol.* 29, 157–168.
- (70) Moul, P. R., and Harvey, J. (2008) Hormonal regulation of hippocampal dendritic morphology and synaptic plasticity. *Cell Adh Migr* 2, 269–275.
- (71) Oomura, Y., Hori, N., Shiraishi, T., Fukunaga, K., Takeda, H., Tsuji, M., Matsumiya, T., Ishibashi, M., Aou, S., Li, X. L., Kohno, D., Uramura, K., Sougawa, H., Yada, T., Wayner, M. J., and Sasaki, K. (2006) Leptin facilitates learning and memory performance and enhances hippocampal CA1 long-term potentiation and CaMK II phosphorylation in rats. *Peptides* 27, 2738–2749.
- (72) Moon, H. S., Dincer, F., and Mantzoros, C. S. (2013) Amylin-induced downregulation of hippocampal neurogenesis is attenuated by leptin in a STAT3/AMPK/ERK-dependent manner in mice. *Diabetologia* 56, 627–634.
- (73) Marwarha, G., and Ghribi, O. (2012) Leptin signaling and Alzheimer's disease. *Am. J. Neurodegener. Dis.* 1, 245–265.
- (74) Balland, E., and Cowley, M. A. (2015) New insights in leptin resistance mechanisms in mice. *Front. Neuroendocrinol.* 39, 59.
- (75) Alzoubi, K. H., Aleisa, A. M., and Alkadhi, K. A. (2005) Impairment of long-term potentiation in the CA1, but not dentate gyrus, of the hippocampus in Obese Zucker rats: role of calcineurin and phosphorylated CaMKII. *J. Mol. Neurosci.* 27, 337–346.
- (76) Fukui, K., Onodera, K., Shinkai, T., Suzuki, S., and Urano, S. (2001) Impairment of learning and memory in rats caused by oxidative stress and aging, and changes in antioxidative defense systems. *Ann. N. Y. Acad. Sci.* 928, 168–175.
- (77) Kumar, J. S., and Menon, V. P. (1993) Effect of diabetes on levels of lipid peroxides and glycolipids in rat brain. *Metab., Clin. Exp.* 42, 1435–1439.
- (78) Moreira, P. I., Santos, M. S., Seica, R., and Oliveira, C. R. (2007) Brain mitochondrial dysfunction as a link between Alzheimer's disease and diabetes. *J. Neurol. Sci.* 257, 206–214.
- (79) Moreira, P. I., Santos, M. S., Sena, C., Seica, R., and Oliveira, C. R. (2005) Insulin protects against amyloid beta-peptide toxicity in brain mitochondria of diabetic rats. *Neurobiol. Dis.* 18, 628–637.
- (80) Moreira, P. I., Santos, M. S., Moreno, A. M., Seica, R., and Oliveira, C. R. (2003) Increased vulnerability of brain mitochondria in diabetic (Goto-Kakizaki) rats with aging and amyloid-beta exposure. *Diabetes* 52, 1449–1456.
- (81) Sima, A. A., Zhang, W., Kreipke, C. W., Rafols, J. A., and Hoffman, W. H. (2009) Inflammation in Diabetic Encephalopathy is Prevented by C-Peptide. *Rev. Diabet Stud* 6, 37–42.
- (82) Kuhad, A., Bishnoi, M., Tiwari, V., and Chopra, K. (2009) Suppression of NF-kappabeta signaling pathway by tocotrienol can prevent diabetes associated cognitive deficits. *Pharmacol., Biochem. Behav.* 92, 251–259.
- (83) Kuhad, A., and Chopra, K. (2008) Effect of sesamol on diabetes-associated cognitive decline in rats. *Exp. Brain Res.* 185, 411–420.
- (84) Arvanitakis, Z., Wilson, R. S., Bienias, J. L., Evans, D. A., and Bennett, D. A. (2004) Diabetes mellitus and risk of Alzheimer disease and decline in cognitive function. *Arch. Neurol.* 61, 661–666.
- (85) Leonard, B. E. (2007) Inflammation, depression and dementia: are they connected? *Neurochem. Res.* 32, 1749–1756.
- (86) Minghetti, L., Ajmone-Cat, M. A., De Berardinis, M. A., and De Simone, R. (2005) Microglial activation in chronic neurodegenerative diseases: roles of apoptotic neurons and chronic stimulation. *Brain Res. Rev.* 48, 251–256.
- (87) Valente, T., Gella, A., Fernandez-Busquets, X., Unzeta, M., and Durany, N. (2010) Immunohistochemical analysis of human brain suggests pathological synergism of Alzheimer's disease and diabetes mellitus. *Neurobiol. Dis.* 37, 67–76.
- (88) Puig, K. L., Floden, A. M., Adhikari, R., Golovko, M. Y., and Combs, C. K. (2012) Amyloid precursor protein and proinflammatory changes are regulated in brain and adipose tissue in a murine model of high fat diet-induced obesity. *PLoS One* 7, e30378.
- (89) Dinel, A. L., Andre, C., Aubert, A., Ferreira, G., Laye, S., and Castanon, N. (2011) Cognitive and emotional alterations are related to hippocampal inflammation in a mouse model of metabolic syndrome. *PLoS One* 6, e24325.

- (90) Nagayach, A., Patro, N., and Patro, I. (2014) Astrocytic and microglial response in experimentally induced diabetic rat brain. *Metab. Brain Dis.* 29, 747–761.
- (91) Miao, Y., He, T., Zhu, Y., Li, W., Wang, B., and Zhong, Y. (2015) Activation of Hippocampal CREB by Rolipram Partially Recovers Balance Between TNF- α and IL-10 Levels and Improves Cognitive Deficits in Diabetic Rats. *Cell. Mol. Neurobiol.* 35, 1157.
- (92) Wang, J., Li, G., Wang, Z., Zhang, X., Yao, L., Wang, F., Liu, S., Yin, J., Ling, E. A., Wang, L., and Hao, A. (2012) High glucose-induced expression of inflammatory cytokines and reactive oxygen species in cultured astrocytes. *Neuroscience* 202, 58.
- (93) Sima, A. A. (2010) Encephalopathies: the emerging diabetic complications. *Acta Diabetol.* 47, 279–293.
- (94) Hernandez-Fonseca, J. P., Rincon, J., Pedraza, A., Viera, N., Arcaya, J. L., Carrizo, E., and Mosquera, J. (2009) Structural and ultrastructural analysis of cerebral cortex, cerebellum, and hypothalamus from diabetic rats. *Exp. Diabetes Res.* 2009, 329632.
- (95) Manschot, S. M., Brands, A. M., van der Grond, J., Kessels, R. P., Algra, A., Kappelle, L. J., and Biessels, G. J. (2006) Brain magnetic resonance imaging correlates of impaired cognition in patients with type 2 diabetes. *Diabetes* 55, 1106–1113.
- (96) Wu, W., Brickman, A. M., Luchsinger, J., Ferrazzano, P., Pichiale, P., Yoshita, M., Brown, T., DeCarli, C., Barnes, C. A., Mayeux, R., Vannucci, S. J., and Small, S. A. (2008) The brain in the age of old: the hippocampal formation is targeted differentially by diseases of late life. *Ann. Neurol.* 64, 698–706.
- (97) Rasgon, N. L., Kenna, H. A., Woolie, T. E., Kelley, R., Silverman, D., Brooks, J., Williams, K. E., Powers, B. N., Hallmayer, J., and Reiss, A. (2009) Insulin resistance and hippocampal volume in women at risk for Alzheimer's disease. *Neurobiol. Aging* 32, 1942.
- (98) Malone, J. I., Hanna, S., Saporta, S., Mervis, R. F., Park, C. R., Chong, L., and Diamond, D. M. (2008) Hyperglycemia not hypoglycemia alters neuronal dendrites and impairs spatial memory. *Pediatr. Diabetes* 9, 531–539.
- (99) Martínez-Tellez, R., Gómez-Villalobos, M. J., and Flores, G. (2005) Alteration in dendritic morphology of cortical neurons in rats with diabetes mellitus induced by streptozotocin. *Brain Res.* 1048, 108–115.
- (100) McEwen, B. S., Magarinos, A. M., and Reagan, L. P. (2002) Studies of hormone action in the hippocampal formation: possible relevance to depression and diabetes. *J. Psychosom. Res.* 53, 883–890.
- (101) Tomassoni, D., Nwankwo, I. E., Gabrielli, M. G., Bhatt, S., Muhammad, A. B., Lokhandwala, M. F., Tayebati, S. K., and Amenta, F. (2013) Astroglialosis in the brain of obese Zucker rat: a model of metabolic syndrome. *Neurosci. Lett.* 543, 136–141.
- (102) Jackson-Guilford, J., Leander, J. D., and Nisenbaum, L. K. (2000) The effect of streptozotocin-induced diabetes on cell proliferation in the rat dentate gyrus. *Neurosci. Lett.* 293, 91–94.
- (103) Alvarez-Nolting, R., Arnal, E., Barcia, J. M., Miranda, M., and Romero, F. J. (2012) Protection by DHA of early hippocampal changes in diabetes: possible role of CREB and NF- κ B. *Neurochem. Res.* 37, 105–115.
- (104) Zhang, W. J., Tan, Y. F., Yue, J. T., Vranic, M., and Wojtowicz, J. M. (2008) Impairment of hippocampal neurogenesis in streptozotocin-treated diabetic rats. *Acta Neurol. Scand.* 117, 205–210.
- (105) Beauquis, J., Roig, P., Homo-Delarche, F., De Nicola, A., and Saravia, F. (2006) Reduced hippocampal neurogenesis and number of hilar neurons in streptozotocin-induced diabetic mice: reversion by antidepressant treatment. *Eur. J. Neurosci* 23, 1539–1546.
- (106) Beauquis, J., Homo-Delarche, F., Giroix, M. H., Ehses, J., Coulaud, J., Roig, P., Portha, B., De Nicola, A. F., and Saravia, F. (2010) Hippocampal neurovascular and hypothalamic-pituitary-adrenal axis alterations in spontaneously type 2 diabetic GK rats. *Exp. Neurol.* 222, 125.
- (107) Sharifi, A. M., Eslami, H., Larijani, B., and Davoodi, J. (2009) Involvement of caspase-8, -9, and -3 in high glucose-induced apoptosis in PC12 cells. *Neurosci. Lett.* 459, 47–51.
- (108) Santiago, A. R., Cristovao, A. J., Santos, P. F., Carvalho, C. M., and Ambrósio, A. F. (2007) High glucose induces caspase-independent cell death in retinal neural cells. *Neurobiol. Dis.* 25, 464–472.
- (109) Liu, D., Zhang, H., Gu, W., and Zhang, M. (2014) Effects of exposure to high glucose on primary cultured hippocampal neurons: involvement of intracellular ROS accumulation. *Neurological sciences: official journal of the Italian Neurological Society and of the Italian Society of Clinical Neurophysiology* 35, 831–837.
- (110) Li, Z. G., Zhang, W., Grunberger, G., and Sima, A. A. (2002) Hippocampal neuronal apoptosis in type 1 diabetes. *Brain Res.* 946, 221–231.
- (111) Sima, A. A., and Li, Z. G. (2005) The effect of C-peptide on cognitive dysfunction and hippocampal apoptosis in type 1 diabetic rats. *Diabetes* 54, 1497–1505.
- (112) Li, Z. G., Zhang, W., and Sima, A. A. (2005) The role of impaired insulin/IGF action in primary diabetic encephalopathy. *Brain Res.* 1037, 12–24.
- (113) Rivera, P., Perez-Martin, M., Pavon, F. J., Serrano, A., Crespillo, A., Cifuentes, M., Lopez-Avalos, M. D., Grondona, J. M., Vida, M., Fernandez-Llebrez, P., de Fonseca, F. R., and Suarez, J. (2013) Pharmacological administration of the isoflavone daidzein enhances cell proliferation and reduces high fat diet-induced apoptosis and gliosis in the rat hippocampus. *PLoS One* 8, e64750.
- (114) Ehrlich, I., Klein, M., Rumpel, S., and Malinow, R. (2007) PSD-95 is required for activity-driven synapse stabilization. *Proc. Natl. Acad. Sci. U. S. A.* 104, 4176–4181.
- (115) Guyot, L. L., Diaz, F. G., O'Regan, M. H., Song, D., and Phillis, J. W. (2001) The effect of streptozotocin-induced diabetes on the release of excitotoxic and other amino acids from the ischemic rat cerebral cortex. *Neurosurgery* 48 (2), 385–391.
- (116) Morris, M. J., and Pavia, J. M. (2004) Increased endogenous noradrenergic and neuropeptide Y release from the hypothalamus of streptozotocin diabetic rats. *Brain Res.* 1006, 100–106.
- (117) Miyata, S., Yamada, N., Hirano, S., Tanaka, S., and Kamei, J. (2007) Diabetes attenuates psychological stress-elicited 5-HT secretion in the prefrontal cortex but not in the amygdala of mice. *Brain Res.* 1147, 233–239.
- (118) Satoh, E., and Takahashi, A. (2008) Experimental diabetes enhances Ca(2+) mobilization and glutamate exocytosis in cerebral synaptosomes from mice. *Diabetes Res. Clin. Pract.* 81, e14.
- (119) Baptista, F. I., Gaspar, J. M., Cristovao, A., Santos, P. F., Kofalvi, A., and Ambrosio, A. F. (2011) Diabetes induces early transient changes in the content of vesicular transporters and no major effects in neurotransmitter release in hippocampus and retina. *Brain Res.* 1383, 257.
- (120) Ramakrishnan, R., Kempuraj, D., Prabhakaran, K., Jayakumar, A. R., Devi, R. S., Suthanthirarajan, N., and Namasivayam, A. (2005) A short-term diabetes induced changes of catecholamines and p38-MAPK in discrete areas of rat brain. *Life Sci.* 77, 1825–1835.
- (121) Yamato, T., Misumi, Y., Yamasaki, S., Kino, M., and Aomine, M. (2004) Diabetes mellitus decreases hippocampal release of neurotransmitters: an in vivo microdialysis study of awake, freely moving rats. *Diabetes, Nutr. Metab.* 17, 128–136.
- (122) Reisi, P., Alaei, H., Babri, S., Sharifi, M. R., Mohaddes, G., and Soleimannejad, E. (2009) Determination of the extracellular basal levels of glutamate and GABA at dentate gyrus of streptozotocin-induced diabetic rats. *Pathophysiology* 16, 63.
- (123) Sherin, A., Anu, J., Peeyush, K. T., Smijin, S., Anitha, M., Roshni, B. T., and Paulose, C. S. (2012) Cholinergic and GABAergic receptor functional deficit in the hippocampus of insulin-induced hypoglycemic and streptozotocin-induced diabetic rats. *Neuroscience* 202, 69–76.
- (124) Ghareeb, D. A., and Hussen, H. M. (2008) Vanadium improves brain acetylcholinesterase activity on early stage alloxan-diabetic rats. *Neurosci. Lett.* 436, 44–47.
- (125) Duarte, J. M., Oses, J. P., Rodrigues, R. J., and Cunha, R. A. (2007) Modification of purinergic signaling in the hippocampus of streptozotocin-induced diabetic rats. *Neuroscience* 149, 382–391.
- (126) Rodrigues, R. J., Alfaro, T. M., Rebola, N., Oliveira, C. R., and Cunha, R. A. (2005) Co-localization and functional interaction

between adenosine A(2A) and metabotropic group 5 receptors in glutamatergic nerve terminals of the rat striatum. *J. Neurochem.* 92, 433–441.

(127) Kloda, A., Clements, J. D., Lewis, R. J., and Adams, D. J. (2004) Adenosine triphosphate acts as both a competitive antagonist and a positive allosteric modulator at recombinant N-methyl-D-aspartate receptors. *Mol. Pharmacol.* 65, 1386–1396.

(128) Biessels, G. J., ter Laak, M. P., Hamers, F. P., and Gispen, W. H. (2002) Neuronal Ca²⁺ dysregulation in diabetes mellitus. *Eur. J. Pharmacol.* 447, 201–209.

(129) Levy, J. (1999) Abnormal cell calcium homeostasis in type 2 diabetes mellitus: a new look on old disease. *Endocr. J.* 10, 1–6.

(130) Levy, J., Gavin, J. R., 3rd, and Sowers, J. R. (1994) Diabetes mellitus: a disease of abnormal cellular calcium metabolism? *Am. J. Med.* 96, 260–273.

(131) Hall, K. E., Liu, J., Sima, A. A., and Wiley, J. W. (2001) Impaired inhibitory G-protein function contributes to increased calcium currents in rats with diabetic neuropathy. *J. Neurophysiol.* 86, 760–770.

(132) Voitenko, N. V., Kruglikov, I. A., Kostyuk, E. P., and Kostyuk, P. G. (1999) Effect of streptozotocin-induced diabetes on the activity of calcium channels in rat dorsal horn neurons. *Neuroscience* 95, 519–524.

(133) Yusaf, S. P., Goodman, J., Gonzalez, I. M., Bramwell, S., Pinnock, R. D., Dixon, A. K., and Lee, K. (2001) Streptozotocin-induced neuropathy is associated with altered expression of voltage-gated calcium channel subunit mRNAs in rat dorsal root ganglion neurones. *Biochem. Biophys. Res. Commun.* 289, 402–406.

(134) Santiago, A. R., Rosa, S. C., Santos, P. F., Cristovao, A. J., Barber, A. J., and Ambrósio, A. F. (2006) Elevated glucose changes the expression of ionotropic glutamate receptor subunits and impairs calcium homeostasis in retinal neural cells. *Invest. Ophthalmol. Visual Sci.* 47, 4130–4137.

(135) Pancani, T., Anderson, K. L., Brewer, L. D., Kadish, I., DeMoll, C., Landfield, P. W., Blalock, E. M., Porter, N. M., and Thibault, O. (2013) Effect of high-fat diet on metabolic indices, cognition, and neuronal physiology in aging F344 rats. *Neurobiol. Aging* 34, 1977–1987.

(136) Biessels, G. J., Kamal, A., Ramakers, G. M., Urban, I. J., Spruijt, B. M., Erkelens, D. W., and Gispen, W. H. (1996) Place learning and hippocampal synaptic plasticity in streptozotocin-induced diabetic rats. *Diabetes* 45, 1259–1266.

(137) Kamal, A., Biessels, G. J., Urban, I. J., and Gispen, W. H. (1999) Hippocampal synaptic plasticity in streptozotocin-diabetic rats: impairment of long-term potentiation and facilitation of long-term depression. *Neuroscience* 90, 737–745.

(138) Di Luca, M., Ruts, L., Gardoni, F., Cattabeni, F., Biessels, G. J., and Gispen, W. H. (1999) NMDA receptor subunits are modified transcriptionally and post-translationally in the brain of streptozotocin-diabetic rats. *Diabetologia* 42, 693–701.

(139) Gardoni, F., Kamal, A., Bellone, C., Biessels, G. J., Ramakers, G. M., Cattabeni, F., Gispen, W. H., and Di Luca, M. (2002) Effects of streptozotocin-diabetes on the hippocampal NMDA receptor complex in rats. *J. Neurochem.* 80, 438–447.

(140) Chabot, C., Massicotte, G., Milot, M., Trudeau, F., and Gagne, J. (1997) Impaired modulation of AMPA receptors by calcium-dependent processes in streptozotocin-induced diabetic rats. *Brain Res.* 768, 249–256.

(141) Gagne, J., Milot, M., Gelin, S., Lahsaini, A., Trudeau, F., Martinoli, M. G., and Massicotte, G. (1997) Binding properties of glutamate receptors in streptozotocin-induced diabetes in rats. *Diabetes* 46, 841–846.

(142) Tuzcu, M., and Baydas, G. (2006) Effect of melatonin and vitamin E on diabetes-induced learning and memory impairment in rats. *Eur. J. Pharmacol.* 537, 106–110.

(143) Kuhad, A., and Chopra, K. (2007) Curcumin attenuates diabetic encephalopathy in rats: behavioral and biochemical evidences. *Eur. J. Pharmacol.* 576, 34–42.

(144) Kamboj, S. S., Vasishta, R. K., and Sandhir, R. (2010) N-acetylcysteine inhibits hyperglycemia-induced oxidative stress and apoptosis markers in diabetic neuropathy. *J. Neurochem.* 112, 77–91.

(145) Ates, O., Yucel, N., Cayli, S. R., Altinoz, E., Yologlu, S., Kocak, A., Cakir, C. O., and Turkoz, Y. (2006) Neuroprotective effect of etomidate in the central nervous system of streptozotocin-induced diabetic rats. *Neurochem. Res.* 31, 777–783.

(146) Magarinos, A. M., Jain, K., Blount, E. D., Reagan, L., Smith, B. H., and McEwen, B. S. (2001) Peritoneal implantation of macro-encapsulated porcine pancreatic islets in diabetic rats ameliorates severe hyperglycemia and prevents retraction and simplification of hippocampal dendrites. *Brain Res.* 902, 282–287.

(147) Coleman, E. S., Dennis, J. C., Braden, T. D., Judd, R. L., and Posner, P. (2010) Insulin treatment prevents diabetes-induced alterations in astrocyte glutamate uptake and GFAP content in rats at 4 and 8 weeks of diabetes duration. *Brain Res.* 1306, 131–141.

(148) Jafari Anarkooli, I., Sankian, M., Vahedi, F., Bonakdaran, S., Varasteh, A. R., and Haghiri, H. (2009) Evaluation of insulin and ascorbic acid effects on expression of Bcl-2 family proteins and caspase-3 activity in hippocampus of STZ-induced diabetic rats. *Cell. Mol. Neurobiol.* 29, 133–140.

(149) Candeias, E. M., Sebastiao, I. C., Cardoso, S. M., Correia, S. C., Carvalho, C. I., Placido, A. I., Santos, M. S., Oliveira, C. R., Moreira, P. I., and Duarte, A. I. (2015) Gut-brain connection: The neuroprotective effects of the anti-diabetic drug liraglutide. *World J. Diabetes* 6, 807–827.

(150) Parthasarathy, V., and Holscher, C. (2013) Chronic treatment with the GLP1 analogue liraglutide increases cell proliferation and differentiation into neurons in an AD mouse model. *PLoS One* 8, e58784.

(151) Duarte, A. I., Candeias, E., Correia, S. C., Santos, R. X., Carvalho, C., Cardoso, S., Placido, A., Santos, M. S., Oliveira, C. R., and Moreira, P. I. (2013) Crosstalk between diabetes and brain: glucagon-like peptide-1 mimetics as a promising therapy against neurodegeneration. *Biochim. Biophys. Acta, Mol. Basis Dis.* 1832, 527–541.

(152) Parthasarathy, V., and Holscher, C. (2013) The type 2 diabetes drug liraglutide reduces chronic inflammation induced by irradiation in the mouse brain. *Eur. J. Pharmacol.* 700, 42–50.

(153) Pipatpiboon, N., Pintana, H., Pratchayasakul, W., Chattipakorn, N., and Chattipakorn, S. C. (2013) DPP4-inhibitor improves neuronal insulin receptor function, brain mitochondrial function and cognitive function in rats with insulin resistance induced by high-fat diet consumption. *Eur. J. Neurosci* 37, 839–849.

(154) Darsalia, V., Ortsater, H., Olverling, A., Darlof, E., Wolbert, P., Nystrom, T., Klein, T., Sjöholm, A., and Patrone, C. (2013) The DPP-4 inhibitor linagliptin counteracts stroke in the normal and diabetic mouse brain: a comparison with glimepiride. *Diabetes* 62, 1289–1296.

(155) Pintana, H., Apaijai, N., Chattipakorn, N., and Chattipakorn, S. C. (2013) DPP-4 inhibitors improve cognition and brain mitochondrial function of insulin-resistant rats. *J. Endocrinol.* 218, 1–11.

(156) Hamilton, A., Patterson, S., Porter, D., Gault, V. A., and Holscher, C. (2011) Novel GLP-1 mimetics developed to treat type 2 diabetes promote progenitor cell proliferation in the brain. *J. Neurosci. Res.* 89, 481–489.

(157) McClean, P. L., Gault, V. A., Harriott, P., and Holscher, C. (2010) Glucagon-like peptide-1 analogues enhance synaptic plasticity in the brain: a link between diabetes and Alzheimer's disease. *Eur. J. Pharmacol.* 630, 158–162.